

Beta-Aminopropionitrile Treatment Effects on MC3T3-E1 Osteoblast Gene Expression and Type I Collagen Production

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INTRODUCTION

Type I Collagen

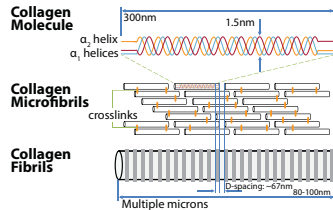
• Cells secrete three alpha helices which form a helical procollagen molecule

• Terminal telopeptide ends of molecules are cleaved by proteinases

• Inline self-assembly of molecules forms microfibrils

• Microfibrils arrange in a quarter-staggered array into fibrils with repeating gap and overlap regions and are stabilized by crosslinks

• Periodicity of the gap and overlap region is referred to as the D-spacing and exists as a distribution of values



• D-spacing provides information on the state and internal structure of tropocollagen as well as post-translational modifications

• Changes in the D-spacing distribution are reflective of disease, tissue type, and drug treatment

Collagen crosslinking

• Crosslinks stabilize collagen molecules within the fibrillar structure and the staggered array

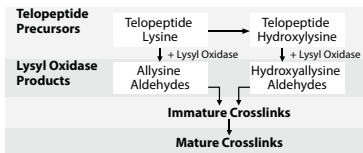
• Enzymatic crosslink formation initiated in telopeptides by lysyl oxidase (LOX) enzyme reaction

Osteolathyrism

• Disease characterized by crosslink deficiency resulting in mechanical defects to bone and connective tissues

• Caused by high dietary consumption of osteolathrogenic compounds such as beta-aminopropionitrile (BAPN)

• BAPN irreversibly binds to the active site of the LOX enzyme, preventing it from acting on telopeptide precursors



Study motivation

• Induce collagen synthesis by osteoblasts and study collagen in its native state

• Research the effects of BAPN on nanoscale collagen morphology and expression of genes relating to collagen synthesis

HYPOTHESIS

BAPN will alter the morphology of type I collagen produced *in vitro* by osteoblasts and drive upregulation of lysyl oxidase to compensate for the reduction in crosslink formation

MATERIALS AND METHODS

Cell culture

• Murine preosteoblasts (MC3T3-E1) cultured in proliferation medium:
 α-MEM medium
 10% fetal bovine serum
 0.05% penicillin/streptomycin
 1% L-glutamine

Collagen synthesis

• MC3T3-E1 cells were cultured in a T75 flask and allowed to proliferate until reaching 80% confluence

Atomic force microscopy

• Cells seeded into 8 dishes, 4 dishes per group (control or BAPN, n=4 each), and differentiated for 2 weeks

• Media was removed and cells were treated with 10 mM EDTA to promote detachment from the extracellular matrix

• Matrix was rinsed with water and air-dried

• Cells were differentiated in proliferation medium supplemented with 50 µg/mL ascorbic acid

• Experimental cultures were supplemented with 0.25 mM BAPN-fumarate

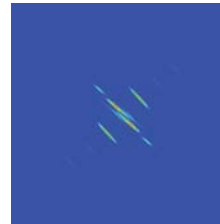
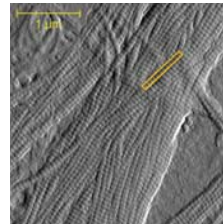
• 500,000 cells were seeded into 60 mm dishes (density: 177 cells/mm²)

• 5 locations per dish were imaged in air by atomic force microscopy

• 3.5 µm x 3.5 µm images from each location

• 2D Fast Fourier Transform (2D-FFT) performed on 10 collagen fibrils per location for D-spacing analysis

• Minimum of 50 fibrils per dish and 200 fibrils per group



Left: AFM error image of collagen with one fibril outlined for 2D-FFT. Right: 2D-FFT corresponding to the outlined fibril.

Quantitative reverse transcription polymerase chain reaction (qRT-PCR)

• Cells seeded into 10 dishes, 5 dishes per group (control or BAPN, n=5 each), and differentiated for 1 week

• SYBR Green primers and master mix used in determining mRNA expression

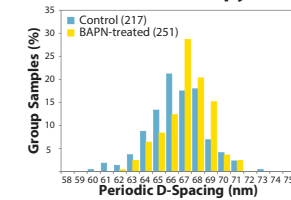
• qRT-PCR performed using an ABI 7500 Fast machine

• Sample/gene combinations run in triplicate with beta-actin as reference gene

• Expression fold change found using Comparative CT method

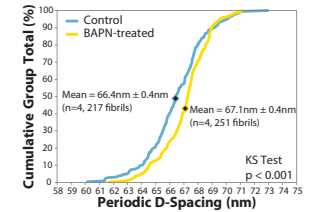
RESULTS AND DISCUSSION

Atomic force microscopy



• Mean D-spacing
 Control: 66.4 nm ± 0.4 nm
 BAPN: 67.1 nm ± 0.4 nm

• Non-parametric U-test revealed p=0.060



• D-spacing distribution range
 Control: 60.2 nm - 72.9 nm
 BAPN: 61.7 nm - 71.1 nm

• Kolmogorov-Smirnov test revealed p<0.001

Quantitative reverse transcription polymerase chain reaction

Target Gene	Fold Change	p-value	
Type I Collagen α ₁	COL1A1	1.10	0.2208
Type I Collagen α ₂	COL1A2	-1.02	0.8133
Lysyl Oxidase	LOX	1.30	0.3610
	COL1A1:COL1A2	-1.13	0.3410

• No significant effect of BAPN treatment on mRNA expression of any of the target genes in BAPN samples relative to controls

Discussion

• Fewer crosslinks are initialized and formed due to BAPN binding to LOX active site

• BAPN caused an increase in the D-spacing distribution of collagen produced *in vitro*

• Crosslinks may compress fibrils driving lower D-spacing in normal collagen

• Fewer crosslinks in the BAPN-treated group could account for the increase in D-spacing

• BAPN treatment of mice *in vivo* causes a decrease in D-spacing distribution in bone (Hammond and Wallace, 2015)

• Post-hoc size analysis of marginally significant mean D-spacing (n=4, p=0.060) indicated a sample size n=6 would be required to detect a statistical significance between groups at 80% power

• Lack of LOX gene upregulation shows no compensatory response to LOX inhibition

• BAPN causes no significant change in expression of genes encoding either a protein helix

CONCLUSION

BAPN alters the nanoscale morphology of type I collagen produced *in vitro* by osteoblasts by causing an upward shift in the D-spacing distribution, but induces no significant change in gene expression relating to collagen synthesis or LOX regulation